



Review Article

Preventive Effect of Antioxidant Vitamins on Cardiovascular Diseases

S. B Mahammad Rahmathulla^{1,2}, Kodidhela Lakshmi Devi^{1,*}

¹ Department of Biochemistry, Sri Krishnadevaraya University, Anantapur, India

² Department of Biochemistry, College of Medicine and Health Sciences, Gondar (Ethiopia).

Received 4th January 2013; Accepted 19th February 2013; Available on line 25th March 2013.

ABSTRACT: Cardiovascular disease (CVD) is the greatest killer in the world today, imposing a significant burden on every nation's health and wealth. Atherosclerosis remains one of the leading reasons for CVD. Subsequent to the discovery that oxidative stress plays a pivotal role in the development and progression of atherosclerosis, vitamins C and E, along with other antioxidants, were studied as potential therapies for the disease. However, while *in vitro* and *in vivo* studies showed promising antiatherogenic effects for vitamins C and E, clinical trials in which patients were given high doses of vitamin C or E showed no benefit and even possible harm. This review will attempt to summarize the known mechanistic data regarding the biochemical effects of vitamins C and E and their relevance to atherosclerosis, and offer an explanation for the failure of clinical trials to show that supplementation with these vitamins provides any benefit when given indiscriminately.

Keywords: Biochemistry; antioxidants; vitamin C; vitamin E; atherosclerosis; CVD

1. INTRODUCTION

Cardiac disorders are of serious medical concern, and are increasing throughout the world. Atherogenesis have provided a growing body of information about molecular mechanisms of plaque growth; however, transition from coronary stability to instability is less well understood due to the lack of animal models reflective of human disease [1a]. Cardiac function is regulated by a complex interplay of neural, hormonal, and mechanical controls. In addition, myocardial function can be altered by free radical generation, an example being the impairment of cardiac function that occurs in the setting of ischemia/reperfusion, a model of acute free radical excess. [1b]. Atherosclerosis remains one of the leading causes of death in Western societies. Interventional therapies have focused on lowering the levels of low density lipoprotein (LDL) cholesterol which is closely correlated with the risk of atherosclerotic vascular disease. Experimental data suggesting that LDL oxidation may be an important process in the development of the atherosclerotic plaque led to the notion that decreasing oxidative stress may help prevent the disease or attenuate its progression [2a]. Various antioxidants, and among them vitamins C and E, have attracted much attention regarding their ability to modulate the progression of atherosclerosis but clinical trials in which high dose supplements of these vitamins have been investigated have shown no benefits, and even possible harm with this intervention. However, *in vitro* and *in vivo* studies showing anti-atherogenic

effects of these antioxidants on cells of the vessel wall raise questions regarding the ability of the trials conducted to reveal their true antiatherogenic potential. This review will give an update on the current knowledge regarding the atheroprotective effects of vitamins C and E, and will attempt to explain the failure of clinical trials to demonstrate benefit from vitamin supplementation.

The Food and Nutrition Board has defined a dietary antioxidant as a substance in commonly consumed foods that significantly decreases the adverse effects of chemically reactive species, such as reactive oxygen and nitrogen species, on normal physiological functions in humans. [2b] Support for the importance of dietary antioxidants in coronary heart disease prevention has come from observational studies, including descriptive, case-control, and cohort studies, in which disease outcomes have been examined in relation to measures of antioxidant intake or tissue levels. [3-5] In many cases, increased antioxidant intake has been shown to be associated with reduced disease risk. This generally has involved increased consumption of antioxidant-rich foods (see Table), although some [6-8] but not all [9] recent results have suggested the possible importance of supplemental levels of antioxidants.

2. Vitamin C (L-Ascorbate)

Vitamin C is known to take part in many physiological processes, and has been proposed to have a beneficial or therapeutic role in immune responses, cardiovascular disease and cancer.

*Corresponding Author:

E-mail address: mdrtl.biochem@gmail.com,

klakshmidevi.bio@gmail.com

Table 1. Vitamin C, E antioxidant vitamins, food sources and their sites of action in cardiomyocytes

S. NO	Name	Food Sources	Site	Action
1	Vitamin C (ascorbic acid)	Fruits (especially citrus) and vegetables, including green and red peppers, tomatoes, potatoes, and green, leafy varieties (eg, spinach and collard greens)	Cytoplasm and plasma	Directly as an antioxidant or as a cofactor for vitamin E
2	Vitamin E (α -tocopherol)	Vegetable oils (eg, soybean, corn, and safflower) and vegetable oil products (eg, margarine), whole grains, wheat germ, nuts and seeds, and green, leafy vegetables	Cytoplasm and plasma	Break lipid peroxidation chain and LDL reaction

Vitamin C is a hydrophilic molecule, and, therefore, it is found mostly in bodily fluids.

Modern food processing methods lead to the loss of vitamin C, as well as many other vitamins and nutrients [10]. In 1928 vitamin C was isolated and recognized as the bioactive molecule that was missing in the diet of sailors, causing scurvy [11].

2.1. L-Ascorbate's structure, Chemistry and Antioxidant Activity

L-Ascorbate's structure that includes two adjacent hydroxyl groups and a carbonyl makes this molecule an excellent hydrogen or electron donor. Therefore, it takes part as a co-factor in many enzymatic reactions, and also acts as a plasma localized anti-oxidant. Once oxidized, ascorbate is turned into ascorbate free radical (AFR), a molecule that is relatively stable due to electron delocalization. Although AFR can donate another electron, it does not undergo further oxidation. Rather, it is reduced back to ascorbate via NADH-dependent and independent mechanisms. AFR accumulation, resulting from increased oxidative conditions, leads to a reaction between two AFR molecules that form one molecule of ascorbate and one molecule of dehydroascorbate (DHA). DHA itself can either be reduced back to ascorbate, or hydrolyzed to gulonic acid [12]. L-Ascorbate fulfills the requirements of an antioxidant, since it can react with radicals and terminate their reaction. Indeed, in the cellular environment where its concentrations are high and recycling mechanisms are abundant, L-ascorbate protects the cell from oxidative stress [13]. L-ascorbate radical can also serve as an electron donor, and actually accelerate redox reactions in the presence of transition metals such as iron or copper. Thus, in the atherosclerotic plaque where ferric iron is present, vitamin C might serve as a pro-oxidant rather than as an anti-oxidant [14].

2.2. L-Ascorbate Metabolism

In truly mammals, L-ascorbate is synthesized endogenously by the enzyme L- γ -gulonolactone oxidase. Nevertheless, in primates and guinea pigs

a functional gene for this enzyme is absent and therefore, the only source of L-ascorbate in guinea pigs and primates is from the diet [12]. Absorption, reabsorption and cellular uptake of L-ascorbate are mediated by the sodium dependent vitamin C transporters (SVCTs). There are two families of these transporters, SVCT1 and SVCT2. Both of these transporters couple the entry of Na⁺ with that of L-ascorbate into cells, against its electrochemical gradient. Different tissues in the body express different types of SVCTs [15]: while both SVCT1 and SVCT2 are expressed in the gastrointestinal tract and mediate absorption of L-ascorbate, only SVCT1 is expressed in the kidney and mediates reabsorption. Thus, knockout of SVCT1 led to renal loss of L-ascorbate in mice and significantly decreased levels of L-ascorbate in plasma [12]. Endothelial cells express SVCT2 alone [16]. The SVCT2 transporter seems to be critical for normal development of blood vessels, as SVCT2^{-/-} mice present with petechia and ecchymoses in the brain shortly after birth [17]. The SVCTs are able to form a gradient of L-ascorbate of up to 1:50. This means that when plasma concentrations are as low as 30-60 μ M, cellular concentration of L-ascorbate can be as high as 4mM, concentrations that are needed for optimal production of Type IV collagen [18].

2.3. Atheroprotective Properties

L-ascorbate not only acts as an antioxidant to reduce vitamin E and also it shows effects on different cells of the vessel wall.

2.3.1. Provocation of Endothelial Cell Proliferation

L-Ascorbate has been shown to promote endothelial cell proliferation, and decrease growth inhibition and apoptosis induced by TNF- α , oxidative stress [19] and oxidized LDL [20]. The proliferative effect of L-ascorbate is thought to be mediated by its effect on Type IV collagen synthesis, which is an integral constituent of the basement membrane and is also responsible for endothelial adhesion. This was proven by showing a decrease in L-ascorbate's proliferative action in

the presence of cis-hydroxypropyl (CHP), which inhibits the enzyme prolyl hydroxylase that is essential for the production of Type IV collagen in endothelial cells. Looking at intracellular signaling, L-ascorbate was shown to decrease p53 levels and increase phosphorylation of the cell cycle regulator Rb, thus rendering it inactive and enabling proliferation.

2.3.2. Inhibition of Endothelial Cell Apoptosis

L-ascorbate is ability to decrease apoptosis in oxidative and inflammatory conditions has been shown to be the result of inhibition of cytochrome C release from mitochondria and prevention of the activation of caspase 9. The same anti-apoptotic effect is exerted by NO as well, and indeed, inhibition of NO production by adding L-NMMA to the culture medium abolished the anti-apoptotic effects of L-ascorbate [21]. Furthermore, other studies have shown that L-ascorbate can potentiate the production of NO by stabilizing tetrahydrobiopterin, the co-factor required for the enzymatic reaction carried out by NOS. This has an effect not only on endothelial survival but also on the entire vessel wall, reducing oxidative stress and inflammation [22].

2.3.3. Augmentation of Endothelial Function

L-ascorbate is expected to reverse endothelial cell dysfunction. Indeed, several independent trials have shown that L-ascorbate effectively reverses endothelial cell dysfunction caused by hypercholesterolemia [23], hypertension, diabetes and atherosclerosis. Reversal of endothelial cell dysfunction was not only shown after acute single intravenous dose of L-ascorbate, but also after chronic (one month) oral intake of the vitamin [24].

2.3.4. Retention of Smooth Muscle Cell Proliferation

L-ascorbate atheroprotective effects are not restricted to the endothelium alone. Aside from the antiatherogenic effect exerted by the overlying endothelium, the smooth muscle cells of the vessel wall are directly affected by L-ascorbate. Most notably, L-ascorbate was shown to decrease smooth muscle cell proliferation in response to mildly oxidized LDL. Furthermore, L-ascorbate was shown to induce smooth muscle cell differentiation in vitro. In vivo studies on rabbits that underwent balloon induced carotid injury showed that not only did L-ascorbate induce differentiation of smooth muscle cells in the neointimal layer of the plaque, but it also prevented dedifferentiation of smooth muscle cells of the media [25]. This effect may have a crucial impact on plaque progression, as dedifferentiated smooth muscle cells cannot only proliferate, but also differentiate to macrophages and thus intensify the inflammation in the vessel wall and accelerate

plaque growth. The prevention of neointimal growth was shown in a clinical trial in which oral intake of L-ascorbate resulted in larger luminal diameter and decreased need for another intervention four months after angioplasty [26].

2.4. L-Ascorbate, Reduce Cardiovascular Events and Overall Mortality

The studies presented suggest that L-ascorbate has an atheroprotective effect, clinical trials regarding its ability to reduce cardiovascular risk and overall mortality have not shown any benefit; L-ascorbate had no added effect in decreasing thickness of the carotid artery wall [27], nor was it able to attenuate coronary atherosclerotic progression [28]. In an eight-year trial that studied the ability of L-ascorbate and vitamin E (either each one alone, or both together) to prevent cardiovascular events, cerebrovascular events and overall mortality of healthy men or men suffering from cardiovascular disease, L-ascorbate did not show any benefit on any of the study endpoints [29].

3. Vitamin E

Vitamin E is mostly found in green vegetables, grains, nuts and various vegetable oils, as well as in eggs and milk. Although it is commonly known today for its antioxidant properties, the first biological role attributed to vitamin E was its necessity for fetal survival [30]. Today vitamin E is known to possess many biological properties, including antioxidant activity and the ability to modulate protein function and gene expression.

3.1. Structure

Vitamin E is most abundant in lipid phase compartments such as the plasma membrane and lipoproteins. It is also found in the membranes of cellular organelles and most notably in the lysosome and the Golgi membrane, where its concentration is more than ten times higher than in other membranes [31]. All vitamin E compounds are lipophilic. The lipophilicity of the compounds is attributed to their hydrophobic tail, a saturated phytyl chain in the tocopherols and an unsaturated phytyl chain in the tocotrienols. The antioxidant activity is attributed to the chromanol group, whose methylation differs among members of the vitamin E group. α -tocopherol, which is the most abundant vitamin E *in vivo*, is methylated on the 5th, 7th and 8th carbon of the chromanol ring [32].

3.2. Metabolism

Vitamin E is absorbed in the gut via micelles, and then incorporated into chylomicrons [33]. When it reaches the circulation, vitamin E is transferred to other lipoproteins by the action of phospholipid transfer protein (PLTP) and to cells by the action of PLTP and lipoprotein lipase (LPL) [34]. Vitamin E is also taken up and re-distributed by the liver, with

the uptake of chylomicrons and the release of very low density lipoproteins (VLDL) [35]. Vitamin E levels are tightly regulated by enzymatic activity of the CYP enzymes, the activity of which changes in response to changes in plasma α -tocopherol levels. Other forms of vitamin E, such as γ -tocopherol, are also metabolized and excreted, but unlike α -tocopherol, they do not have a profound effect on CYP activity. α -Tocopherol can also be excreted in the bile via the Multi Drug Resistance (MDR) family of transporters [36]. The liver can also secrete vitamin E by the α -tocopherol-transfer protein (α -TTP), which is highly specific for α -tocopherol and mediates its transfer to various lipoproteins [37].

3.3. Vitamin E in Relation to Atherosclerosis

Vitamin E identified to be Regulated of Cell Survival, Proliferation and Apoptosis, an inhibitory effect on protein kinase C (PKC) non-antioxidant functions [38]. Vitamin E was found to activate phospho-serine/Threonine phosphatase 2A (PP2A), which is responsible for the dephosphorylation of PKC, a process that occurs on the cell membrane [39]. Vitamin E was also shown to inhibit Protein Kinase B (PKB) and activate protein tyrosine phosphatase, altering cell proliferation and survival, the most prominent effect of vitamin E mediated by PKC inhibition is the reduction of cell proliferation. This has been shown to occur in various cells [40], the inhibition of vascular smooth muscle cells (VSMCs) being most relevant to the attenuation of the atherosclerotic process [41]. Another signaling pathway which is subjected to modulation by vitamin E is the mitogen-activated protein kinase (MAPK) pathway. In VSMCs stimulated by oxidized LDL, vitamin E was shown to decrease MAPK activity and enhance cell survival [42].

3.3.2. Enrichment of Endothelial Function

Vitamin E was shown to enhance various functions of the endothelium, including nitric oxide (NO) release, anti-thrombotic properties and vasodilation. As opposed to the inhibitory effect on arachidonic acid (AA) release and metabolism in other cells, most notably in macrophages, vitamin E leads to an increase in AA release by phospholipase A2 (PLA2) in endothelial cells. Although this effect is accompanied by a decrease in cyclo-oxygenase (COX) 1 and 2 activity, the net effect is an increase in the production of vasodilating prostanoids PGE2 and PGI2 [43]. Additionally, vitamin E was shown to enhance the phosphorylation of endothelial nitric oxide synthase (eNOS) on serine 1177, resulting in an amplification of its action [44–45]. These results translate to increased levels of NO metabolites following vitamin E treatment [46]. However, the effect of vitamin E treatment on endothelial

function is still unclear, as some studies have found an improvement in endothelial function following vitamin E treatment [47-50].

3.3.3. Control of Inflammatory Processes

Vitamin E has been shown to inhibit several inflammatory processes which are known to take place during atherogenesis. Vitamin E inhibits the cellular adhesion process and decreases the expression of various adhesion molecules and chemokines by endothelial cells and leukocytes, both *in vitro*, in a response to a variety of noxious stimuli [51-55], and *in vivo* [56]. Humans receiving high dose vitamin E supplementation demonstrate a decrease in soluble adhesion molecules [57-58]. Additionally, vitamin E was shown to suppress the secretion of pro-inflammatory cytokines such as tumor necrosis factor- α (TNF- α) [59] and interleukin-1 β (IL-1 β) [60- 61]. Scavenger receptors, such as CD36, known to be important for oxidized LDL uptake by macrophages, are down regulated by vitamin E [62- 63]. Finally, vitamin E inhibits the activity of inducible NOS (iNOS) and NADPH oxidase, thereby inhibiting the macrophage respiratory burst [64- 65].

3.3.4. Antioxidant role

Vitamin E is classified as an antioxidant due to its ability to scavenge lipid radicals and terminate oxidative chain reactions. It can terminate radical chain reactions by interacting with the lipid peroxy radical, preventing it from generating a new radical and perpetuating the chain reaction by oxidizing other lipids. This is due to the rate constant of the reaction between lipids and lipid peroxy radicals, which are 1,000-fold, lower than the rate constant of the reaction between α -tocopherol and lipid peroxy radicals (102 M-1S-1 compared to 105–106 M-1S-1). It is unlikely that vitamin E would interfere with the radical chain reaction in other stages. The radical chain reaction is usually initiated by water soluble molecules, where vitamin E is sparse due to its lipophilic nature. Its interaction with lipid radicals is unlikely since the rate constant of the reaction between lipid radicals and oxygen is 100-1,000-fold higher compared to that of lipid radicals and vitamin E. Following its oxidation, vitamin E can be recycled back to its native unoxidized form by various soluble antioxidants such as vitamin C and ubiquinol. This process prevents the accumulation of vitamin E radicals and their subsequent peroxidation of lipids [66], and is considered by some to be critical for the antioxidant activity of vitamin E [67]. It has been suggested that all of the other biological functions of vitamin E are actually a result of its antioxidant activity [68].

3.4. Clinical Studies of Vitamin E

While *in vitro* and *in vivo* studies demonstrate a wide variety of anti-atherogenic effects for vitamin E, these were not translated into the clinical setting as large trials showed no beneficial effect for vitamin E supplementation [69–71]. Overall, supplementation of vitamin E has been shown to cause an increase in mortality in a large meta-analysis [72]. One reason for the failure to show a beneficial effect for vitamin E supplementation may be the fact that it was indiscriminately given to a large population. However, as is true for any pharmaceutical agent, vitamin E supplementation would be predicted to show benefit only in those individuals in which it is needed. Demonstrating this important concept of proper patient selection, when vitamin E was selectively given to DM patients with the Hp 2-2 phenotype, it appeared to provide a significant positive effect on CVD and overall mortality [73]. Additionally, a reanalysis of large clinical trials according to Hp phenotype has shown similar results [74-75].

4. CONCLUSION

The studies presented here have emphasized the marked disparity between the anti-atherogenic effect of the antioxidant vitamins C and E shown in preclinical studies, and their inability to show beneficial effects in clinical trials. *in vitro* and *in vivo* studies may perhaps not accurately represent the biological processes in the human body. However, other explanations may exist for the discrepancy, pertaining to the methodology of the clinical trials. We have presented one example, involving the Hp genotype and vitamin E, wherein only a subset of patients may actually benefit from antioxidant vitamin E therapy. An additional problem with trials regarding antioxidant supplementation is that the timing of their administration may be critical. Vitamin C has been shown to have beneficial effects on processes that occur in the early stages of atherosclerosis; it may prevent lesion formation in the first place or initial plaque growth by improving endothelial function and preventing the formation of the neointima. However, once the atherosclerotic plaque is already formed, the contribution of vitamin C to antiatherogenic processes may be negligible. Most clinical trials were conducted on patients that already suffered from vascular disease. Even among patients without a history of prior symptomatic CVD, the participants' age in most of these studies was over 50, an age in which atherosclerotic lesions and plaques are certainly already present. Trials enrolling younger participants are needed to examine whether vitamin C has an effect in the early stages of atherosclerosis. The dose and mixture of antioxidants that are given may also be critical. In the initial observational dietary studies that demonstrated strong apparent benefit from

antioxidant vitamins, vitamins were obtained from fruits and vegetables. Naturally occurring antioxidant vitamins differ in their formulation (i.e., synthetic vitamin E contains a mixture of stereoisomers while natural vitamin E contains only one stereoisomer) and in the relative concentrations of related molecules. The difference between the many different forms of vitamin E which occur in natural food substances and those that were used in failed clinical trials is striking. Why might antioxidant supplementation be harmful for some populations? Oxidative processes are vital for normal cellular function, and have a pivotal role in various physiological systems, including normal vascular physiology. When given in pharmacological doses, which are much higher than doses that can be attained by dietary intake, antioxidants may attenuate both deleterious and beneficial oxidative processes. This may be the reason why clinical trials that use pharmacological doses of antioxidants do not show a beneficial effect on disease progression when given indiscriminately to all individuals regardless of their baseline level of oxidative stress, more research is needed in this area.

Acknowledgements

Authors would like to thank Department of Biochemistry S.K. University, Anantapuram, A.P, India, Department of Biochemistry, College Of Medicine And Health Sciences- Gondar (Ethiopia).

5. REFERENCES

- [1]. 1a. P. Libby, P.M. Ridker, G.K. Hansson, (2011) Progress and challenges in translating the biology of atherosclerosis. *Nature*, **473**, 317-25., [1b]. R. Bolli, M. Zughuib, X.Y. Li, (1995) Recurrent ischemia in the canine heart causes recurrent bursts of free radical production that have a cumulative effect on contractile function: a pathophysiological basis for chronic myocardial "stunning." *Journal of Clinical Investigation*, **96**, 1066-1084.
- [2]. C.K. Glass, J.L. Witztum, (2001) Atherosclerosis: The road ahead. *Cell*, **104**, 503-516.
- [3]. J.M. Gaziano, J.E. Manson, J.E. Buring, C.H. Hennekens. (1992) Dietary antioxidants and cardiovascular disease. *Annals of the New York Academy of Sciences*, **669**, 249-258.
- [4]. D.L. Tribble, E. Frank (1994) Dietary antioxidants, cancer, and atherosclerotic vascular disease. *Western Journal of Medicine*, **161**, 605– 612.
- [5]. C.H. Hennekens, J.M. Gaziano, J.E. Manson, J.E. Buring (1995) Antioxidant vitamin-cardiovascular disease hypothesis is still promising, but still unproven: the need for

- randomized trials. *American Journal of Clinical Nutrition*, **62**,1377S-1380S.
- [6]. M.J. Stampfer, C.H. Hennekens, J.E. Manson, G.A. Colditz, B. Rosner, W.C. Willett. (1993) Vitamin E consumption and the risk of coronary heart disease in women. *New England Journal of Medicine*, **328**,1444 - 1449.
- [7]. E.B. Rimm, M.J. Stampfer, A. Ascherio, E. Giovannucci, G.A. Colditz, W.C. Willett (1993) Vitamin E consumption and the risk of coronary heart disease in men. *New England Journal of Medicine*, **328**,1450-1456.
- [8]. J.E. Enstrom, L.E. Kanim, M.A. Klein. (1992) Vitamin C intake and mortality among a sample of the United States population. *Epidemiology*, **3**, 194-202.
- [9]. L.H. Kushi, A.R. Folsom, R.J. Prineas, P.J. Mink, Y. Wu, R.M. Bostick. (1996) Dietary antioxidant vitamins and death from coronary heart disease in postmenopausal women *New England Journal of Medicine*, **334**, 1156-1162.
- [10]. F.Deruelle, B. Baron, (2008) Vitamin C: is supplementation necessary for optimal health? *Journal of Alternative and Complementary Medicine*, **14**, 1291-1298.
- [11]. J.H. Baron, (2009) Sailors' scurvy before and after James Lind-a reassessment. *Nutrition Reviews*, **67**, 315-332.
- [12]. R.Stocker, Jr. J.F. Keaney, (2005) New insights on oxidative stress in the artery wall. *Journal of Thrombosis and Haemostasis*, **3**, 1825-1834.
- [13]. R. Aguirre, J.M. May, (2008) Inflammation in the vascular bed: importance of vitamin C. *Pharmacology & Therapeutics*, **119**, 96-103.
- [14]. A.Carr, B. Frei, (1999) Does vitamin C act as a pro-oxidant under physiological conditions? *FASEB Journal*, **13**, 1007-1024.
- [15]. H.Tsukaguchi, T.Tokui, B. Mackenzie, U.V. Berger, X.Z. Chen, Y. Wang, R.F. Brubaker, M.A. Hediger, (1999) A family of mammalian Na⁺-dependent L-ascorbic acid transporters. *Nature*, **399**, 70-75.
- [16]. T. Seno, N. Inoue, K. Matsui, J. Ejiri, K. Hirata, S. Kawashima, M.Yokoyama, (2004) Functional expression of sodium-dependent vitamin C transporter 2 in human endothelial cells. *Journal of Vascular Research*, **41**, 345-351.
- [17]. S.Sotiriou, S. Gispert, J. Cheng, Y. Wang, A.Chen, S. Hoogstraten-Miller, G.F. Miller, O. Kwon, M. Levine, S.H. Guttentag, R.L. Nussbaum, (2002) Ascorbic-acid transporter Slc23a1 is essential for vitamin C transport into the brain and for perinatal survival. *Nature Medicine*, **8**, 514-517.
- [18]. J.M. May, Z.C. Qu, (2005) Transport and intracellular accumulation of vitamin C in endothelial cells: relevance to collagen synthesis. *Archives of Biochemistry and Biophysics*, **434**, 178-186.
- [19]. R.W. Saeed, T. Peng, C.N. Metz, (2003) Ascorbic acid blocks the growth inhibitory effect of tumor necrosis factor-alpha on endothelial cells. *Experimental Biology and Medicine (Maywood)*, **228**, 855-865.
- [20]. G.Ulrich-Merzenich, C. Metzner, B. Schiermeyer, H.Vetter, (2002) Vitamin C and vitamin E antagonistically modulate human vascular endothelial and smooth muscle cell DNA synthesis and proliferation. *European Journal of Nutrition*, **41**, 27-34.
- [21]. L.Rossig, J. Hoffmann, B. Hugel, Z. Mallat, A. Haase, J.M. Freyssinet, A.Tedgui, A. Aicher, A.M. Zeiher, S. Dimmeler, (2001) Vitamin C inhibits endothelial cell apoptosis in congestive heart failure. *Circulation*, **104**, 2182-2187.
- [22]. R. Heller, A. Unbehaun, B. Schellenberg, B. Mayer, G. Werner-Felmayer, E.R. Werner, (2001), L-ascorbic acid potentiates endothelial nitric oxide synthesis via a chemical stabilization of tetrahydrobiopterin. *Journal of Biological Chemistry*, **276**, 40-47.
- [23]. H.H. Ting, F.K. Timimi, E.A. Haley, M.A. Roddy, P. Ganz, M.A. Creager, (1997) Vitamin C improves endothelium-dependent vasodilation in forearm resistance vessels of humans with hypercholesterolemia. *Circulation*, **95**, 2617-2622.
- [24]. N.Gokce, Jr J.F, Keaney, B. Frei, M. Holbrook, M. Olesiak, B.J. Zachariah, C. Leeuwenburgh, J.W. Heinecke, J.A. Vita, (1999), Long-term ascorbic acid administration reverses endothelial vasomotor dysfunction in patients with coronary artery disease. *Circulation*, **99**, 3234-3240.
- [25]. E. Arakawa, K. Hasegawa, J. Irie, S. Ide, J. Ushiki, K. Yamaguchi, S. Oda, Y. Matsuda, (2003) L-ascorbic acid stimulates expression of smooth muscle-specific markers in smooth muscle cells both in vitro and in vivo. *Journal of Cardiovascular Pharmacology*, **2**, 745-751.
- [26]. H.Tomoda, M.Yoshitake, K. Morimoto, N. Aoki, (1996) Possible prevention of postangioplasty restenosis by ascorbic acid. *American Journal of Cardiology*, **78**, 1284-1286.
- [27]. S.P. Azen, D.Qian, W.J. Mack, A. Sevanian, R.H. Selzer, C.R. Liu, C.H. Liu, H.N. Hodis, (1996) Effect of supplementary antioxidant vitamin intake on carotid arterial wall intima-media thickness in a controlled clinical trial of cholesterol lowering. *Circulation*, **94**, 2369-2372.
- [28]. H.N. Hodis, W.J. Mack, L. LaBree, L. Cashin-Hemphill, A. Sevanian, R. Johnson,

- S.P. Azen, (1995) Serial coronary angiographic evidence that antioxidant vitamin intake reduces progression of coronary artery atherosclerosis. *JAMA*, **273**, 1849-1854.
- [29]. H.D. Sesso, J.E. Buring, W.G. Christen, T. Kurth, C. Belanger, J. MacFadyen, V. Bubes, J.E. Manson, R.J. Glynn, J.M. Gaziano, (2008), Vitamins E and C in the prevention of cardiovascular disease in men: the Physicians' Health Study II randomized controlled trial. *JAMA*, **300**, 2123-2133.
- [30]. H.M. Evans, K.S. Bishop, (1922) On the existence of a hitherto unrecognized dietary factor essential for reproduction. *Science*, **56**, 650-651.
- [31]. Y. Zhang, M. Turunen, E.L. Appelkvist, (1996) Restricted uptake of dietary coenzyme Q is in contrast to the unrestricted uptake of alpha-tocopherol into rat organs and cells. *Journal of Nutrition*, **126**, 2089-2097.
- [32]. E. Niki, (1987) Antioxidants in relation to lipid peroxidation. *Chemistry and Physics of Lipids*, **44**, 227-253.
- [33]. R.J. Sokol, J.E. Heubi, S. Iannaccone, K.E. Bove, W.F. Balistreri, (1983) Mechanism causing vitamin E deficiency during chronic childhood cholestasis. *Gastroenterology*, **85**, 1172-1182.
- [34]. G.M. Kostner, K. Oettl, M. Jauhiainen, C. Ehnholm, H. Esterbauer, H. Dieplinger, (1995) Human plasma phospholipid transfer protein accelerates exchange/transfer of alpha-tocopherol between lipoproteins and cells. *Biochemical Journal*, **305**, 659-667.
- [35]. R. Havel, (1994) McCollum Award Lecture, 1993: Triglyceride-rich lipoproteins and atherosclerosis-new perspectives. *American Journal of Clinical Nutrition*, **59**, 795-796.
- [36]. D.J. Mustacich R.S., Bruno, M.G. Traber, (2007) Vitamin E. *Vitamins & Hormones*, **76**, 1-21.
- [37]. R. Brigelius-Flohe, M.G. Traber, (1999) Vitamin E: Function and metabolism. *FASEB Journal*, **13**, 1145-1155.
- [38]. C.W. Mahoney, A. Azzi, (1988) Vitamin E inhibits protein kinase C activity. *Biochemical and Biophysical Research Communications*, **154**, 694-697.
- [39]. R.T. Boudreau, R. Garduno, T.J. Lin, (2002) Protein phosphatase 2A and protein kinase Calpha are physically associated and are involved in Pseudomonas aeruginosa-induced interleukin 6 production by mast cells. *Journal of Biological Chemistry*, **277**, 5322-5329.
- [40]. J.M. Zingg, (2007) Modulation of signal transduction by vitamin E. *Molecular Aspects of Medicine*, **28**, 481-506.
- [41]. A. Tasinato, D. Boscoboinik, G.M. Bartoli, P. Maroni, A. Azzi, (1995) d-alpha-tocopherol inhibition of vascular smooth muscle cell proliferation occurs at physiological concentrations, correlates with protein kinase C inhibition, and is independent of its antioxidant properties. *Proc. Nat. Acad. Sci. USA*, **92**, 12190-12194.
- [42]. F. de Nigris, F. Franconi, I. Maida, G. Palumbo, V. Anania, C. Napoli, (2000) Modulation by alpha and gamma-tocopherol and oxidized low-density lipoprotein of apoptotic signaling in human coronary smooth muscle cells. *Biochemical Pharmacology*, **59**, 1477-1487.
- [43]. D. Wu, L. Liu, M. Meydani, S.N. Meydani, (2004) Effect of vitamin E on prostacyclin (PGI₂) and prostaglandin (PG) E₂ production by human aorta endothelial cells: mechanism of action. *Annals of the New York Academy of Sciences*, **1031**, 425-427.
- [44]. R. Heller, M. Hecker, N. Stahmann, J.J. Thiele, G. Werner-Felmayer, E.R. Werner, (2004) Alpha-tocopherol amplifies phosphorylation of endothelial nitric oxide synthase at serine 1177 and its short-chain derivative trolox stabilizes tetrahydrobiopterin. *Free Radical Biology & Medicine*, **37**, 620-631.
- [45]. R. Heller, G. Werner-Felmayer, E.R. Werner, (2004) Alpha-Tocopherol and endothelial nitric oxide synthesis. *Annals of the New York Academy of Sciences*, **1031**, 74-85.
- [46]. G. Desideri, M.C. Marinucci, G. Tomassoni, P.G. Masci, A. Santucci, C. Ferri, (2002) Vitamin E supplementation reduces plasma vascular cell adhesion molecule-1 and von Willebrand factor levels and increases nitric oxide concentrations in hypercholesterolemic patients. *Journal of Clinical Endocrinology & Metabolism*, **87**, 2940-2945.
- [47]. D. Green, G. O'Driscoll, J.M. Rankin, A.J. Maiorana, R.R. Taylor, (1998) Beneficial effect of vitamin E administration on nitric oxide function in subjects with hypercholesterolaemia. *Clinical Science (Lond)*, **95**, 361-367.
- [48]. T. Motoyama, H. Kawano, K. Kugiyama, O. Hirashima, M. Ohgushi, R. Tsunoda, Y. Moriyama, Y. Miyao, M. Yoshimura, H. Ogawa, H. Yasue, (1998) Vitamin E administration improves impairment of endothelium-dependent vasodilation. *Journal of American College of Cardiology Foundation*, **32**, 1672-1679.]
- [49]. T. Neunteufl, K. Kostner, R. Katzenschlager, M. Zehetgruber, G. Maurer, F. Weidinger, (1998) Additional benefit of vitamin E supplementation to simvastatin therapy on vasoreactivity of the brachial artery of

- hypercholesterolemic men, *Journal of American College of Cardiology Foundation*, **32**, 711-716.
- [50]. T. Heitzer, S. Yla Herttuala, E. Wild, J. Luoma, H. Drexler, (1999) Effect of vitamin E on endothelial vasodilator function in patients with hypercholesterolemia, chronic smoking or both. *Journal of American College of Cardiology Foundation*, **33**, 499-505.
- [51]. T. Yoshikawa, N. Yoshida, H. Manabe, Y. Terasawa, T. Takemura, M. Kondo, (1998) Alpha tocopherol protects against expression of adhesion molecules on neutrophils and endothelial cells. *Bio Factors*, **7**, 15-19.
- [52]. R. Faruqi, C. de la Motte, P.E. DiCorleto, (1994) Alpha-tocopherol inhibits agonist-induced monocytic cell adhesion to cultured human endothelial cells. *Journal of Clinical Investigation*, **94**, 592-600.
- [53]. N. Yoshida, T. Yoshikawa, H. Manabe, Y. Terasawa, M. Kondo, N. Noguchi, E.Niki, (1999) Vitamin E protects against polymorphonuclear leukocyte-dependent adhesion to endothelial cells. *Journal of Leukocyte Biology*, **65**, 757-763.
- [54]. N. Noguchi, R. Hanyu, A. Nonaka, Y. Okimoto, T. Kodama, (2003) Inhibition of THP-1 cell adhesion to endothelial cells by alpha-tocopherol and alpha-tocotrienol is dependent on intracellular concentration of the antioxidants. *Free Radical Biology & Medicine*, **34**, 1614-1620.
- [55]. D.Wu, T. Koga, K.R. Martin, M. Meydani, (1999) Effect of vitamin E on human aortic endothelial cell production of chemokines and adhesion to monocytes. *Atherosclerosis*, **147**, 297-307.
- [56]. T. Koga, P. Kwan, L. Zubik, C. Ameho, D. Smith, M. Meydani, (2004) Vitamin E supplementation suppresses macrophage accumulation and endothelial cell expression of adhesion molecules in the aorta of hypercholesterolemic rabbits. *Atherosclerosis*, **176**, 265-272.
- [57]. G. Desideri, G. Croce, M.C. Marinucci, P.G. Masci, M. Stati, L.Valeri, A. Santucci, C. Ferri, (2002) Prolonged, low dose alpha-tocopherol therapy counteracts intercellular cell adhesion molecule- 1 activation. *Clinica Chimica Acta*, **320**, 5-9.
- [58]. B. van Dam, V.W. van Hinsbergh, C.D. Stehouwer, A. Versteilen, H. Dekker, R. Buytenhek, H.M. Princen, C.G. Schalkwijk, (2003) Vitamin E inhibits lipid peroxidation-induced adhesion molecule expression in endothelial cells and decreases soluble cell adhesion molecules in healthy subjects. *Cardiovascular Research*, **57**, 563-571.
- [59]. S. Devaraj, I. Jialal, (2005) Alpha-tocopherol decreases tumor necrosis factor-alpha mRNA and protein from activated human monocytes by inhibition of 5-lipoxygenase. *Free Radical Biology & Medicine*, **38**, 1212-1220.
- [60]. S. Devaraj, I. Jialal, (1999) Alpha-tocopherol decreases interleukin-1 beta release from activated human monocytes by inhibition of 5-lipoxygenase. *Arteriosclerosis, Thrombosis, and Vascular Biology*, **19**, 1125-1133.
- [61]. S. Devaraj, I. Jialal, (1996) The effects of alpha tocopherol supplementation on monocyte function. Decreased lipid oxidation, interleukin 1 beta secretion, and monocyte adhesion to endothelium. *Journal of Clinical Investigation*, **98**, 756-763.
- [62]. A. Munteanu, J.M Zingg, A. Azzi, (2004) Anti-atherosclerotic effects of vitamin E--myth or reality? *Journal of Cellular and Molecular Medicine*, **8**, 59-76.
- [63]. A. Munteanu, M. Taddei, I. Tamburini, E. Bergamini, A. Azzi, J.M. Zingg, (2006) Antagonistic effects of oxidized low density lipoprotein and alpha-tocopherol on CD36 scavenger receptor expression in monocytes: involvement of protein kinase B and peroxisome proliferator-activated receptor-gamma. *Journal of Biological Chemistry*, **281**, 6489-6497.
- [64]. K.L. Khanduja, P.K. Avti, S. Kumar, V. Pathania, C.M. Pathak, (2005) Inhibitory effect of vitamin E on proinflammatory cytokines-and endotoxin-induced nitric oxide release in alveolar macrophages. *Life Sciences*, **76**, 2669-2680.
- [65]. O. Cachia, J.E. Benna, E. Pedruzzi, B. Descomps, M.A. Gougerot-Pocidalo, C.L.Leger, (1998) alpha-tocopherol inhibits the respiratory burst in human monocytes. Attenuation of p47 (phox) membrane translocation and phosphorylation. *Journal of Biological Chemistry*, **273**, 32801-32805.
- [66]. R. Brigelius-Flohe, (2009) Vitamin E: the shrew waiting to be tamed. *Free Radic. Biol. Med.*, **46**, 543-554.
- [67]. A.C. Carr, B.Z. Zhu B. Frei, (2000) Potential antiatherogenic mechanisms of ascorbate (vitamin C) and alpha-tocopherol (vitamin E). *Circulation Research*, **87**, 349-354.
- [68]. M.G. Traber, J. Atkinson, (2007) Vitamin E, antioxidant and nothing more *Free Radical Biology & Medicine*, **43**, 4-15.
- [69]. E. Lonn, J. Bosch, S. Yusuf, P. Sheridan, J. Pogue, J.M, Arnold, C. Ross, A. Arnold, P. Sleight, J. Probstfield, G.R. Dagenais, (2005) Effects of long-term vitamin E supplementation on cardiovascular events and cancer: a randomized controlled trial. *JAMA*, **293**, 1338-1347.

- [70]. I.M. Lee, N.R. Cook, J.M. Gaziano, D. Gordon, P.M. Ridker, J.E. Manson, C.H. Hennekens, J.E. Buring, (2005) Vitamin E in the primary prevention of cardiovascular disease and cancer: the Women's Health Study: a randomized controlled trial. *JAMA*, **294**, 56-65.
- [71]. M.K. Kataja-Tuomola, J.P. Kontto, S. Mannisto, D. Albanes, J.R. Virtamo, (2010) Effect of alphas-tocopherol and beta-carotene supplementation on macrovascular complications and total mortality from diabetes: results of the ATBC Study. *Annals of Medicine*, **42**, 178-186.
- [72]. G. Bjelakovic, D. Nikolova, L.L. Gluud, R.G. Simonetti, C. Gluud, (2007) Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA*, **297**, 842-857.
- [73]. U. Milman, S. Blum, C. Shapira, D. Aronson, R. Miller-Lotan, Y. Anbinder, J. Alshiek, L. Bennett, M. Kostenko, M. Landau, S. Keidar, Y. Levy, A. Khemlin, A. Radan, A.P. Levy, (2008) Vitamin E supplementation reduces cardiovascular events in a subgroup of middle-aged individuals with both type 2 diabetes mellitus and the haptoglobin 2-2 genotype: a prospective double-blinded clinical trial. *Arteriosclerosis, Thrombosis, and Vascular Biology*, **28**, 341-347.
- [74]. A.P. Levy, H.C. Gerstein, R. Miller-Lotan, R. Ratner, M. McQueen, E. Lonn, J. Pogue, (2004) The effect of vitamin E supplementation on cardiovascular risk in diabetic individuals with different haptoglobin phenotypes. *Diabetes Care*, **27**, 2767.
- [75]. S.Blum, M.Vardi, N.S. Levy, R. Miller-Lotan, A.P. Levy, (2010) The effect of vitamin E supplementation on cardiovascular risk in diabetic individuals with different haptoglobin phenotypes. *Atherosclerosis*, **211**, 25-27.

***Biographical Sketch**



Dr. Kodihela Lakshmi Devi did her B. Sc at Sri Venkateswara University, A.P, India. Dr. K. Lakshmi Devi had completed M.Sc. and Ph.D, at Sri Krishnadevaraya University. Her major research area is cardiovascular diseases in Clinical Biochemistry. She joined as Assistant professor at Department of Biochemistry in 1989; she worked as head and as well as chairperson B. O. S. At present she is working as Professor of Biochemistry, Sri Krishnadevaraya University, Anantapur, India.